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DISTINCT ROLES OF GAMMA-AMINOBUTYRIC ACID TYPE A RECEPTOR SUBTYPES: A FOCUS ON PHASIC AND TONIC INHIBITION

GAMA AMİNOBÜTİRİK ASİT TİP A RESEPTÖR ALT TİPLERİNİN FARKLILAŞMIŞ FONKSİYONLARI: FAZİK VE TONİK İNHİBASYON

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Abstract

The gamma-aminobutyric acid type A receptors (GABA_ARs), belonging to the superfamily of Cys-loop receptors, responsible for the inhibitory transmission in the vertebrate central nervous system. Assembled from a pool of 19 subunits, the subunit composition of heteropentameric GABA_ARs impacts on receptor's function, physiology, cellular and subcellular localization in the cell membrane, i.e., synaptic or extrasynaptic. γ 2 containing GABA_ARs (γ 2-GABA_ARs) are clustered in the synapses and mediate classical fast synaptic inhibition called phasic inhibition. δ subunit containing GABA_ARs (δ -GABA_ARs) are located extrasynaptically and mediate a different form of inhibition called tonic inhibition critical for the threshold of action potential generation and neuronal excitability. Thus, distinct physiological roles of synaptic and extrasynaptic GABA_AR subtypes lead to the question to ask about the possibility of subtype selective drugs. In the light of accumulating data from X-ray crystal structures of vertebrate, invertebrate and prokaryotic Cys-loop receptor family members, new opportunities arise for the development of novel drugs targeting specifically these subtypes of GABA_ARs for the treatment of various neuropathological conditions.

Keywords: GABAA Receptors, GABA, phasic inhibition, tonic inhibition, gamma2 (γ 2) subunit, delta (δ) subunit, Cys-loop receptors, synaptic, extrasynaptic

Özet

Memelilerde beyinde inhibasyonun iletiminden sorumlu birincil reseptör olan gama amino butirik asit tip A reseptörleri (GABA_ARs), Cys-loop reseptörleri familyasına bağlıdır. 19 alt üniteden oluşan bir havuzdan belirli kombinasyonlarla pentamer olarak organize olan bu reseptörlerin kompozisyonu, reseptörün fonksiyonu, fizyolojisi, bulunduğu hücre tipi ve sinaptik ya da ekstra-sinaptik gibi hücre zarında belirli bir lokalizasyonda bulunmasına etki eder. γ 2 alt ünitesini bulunduran GABA_AR alt tipi (γ 2-GABA_ARs) sinaptik olarak lokalize olup fazik inhibasyon olarak bilinen tipik hızlı inhibasyonun iletiminden sorumludurlar. δ alt ünitesini bulunduran GABA_ARs (δ -GABA_ARs) ekstrasinaptik olarak lokalize olup fazik inhibasyondan daha farklı bir inhibasyon türü olan ve aksiyon potansiyelinin oluşması için gereken eşik değerinde ve hücre uyarılabilirliğinde rol oynayan tonik inhibasyonun iletiminden sorumludurlar. Sinaptik ve ekstrasinaptik GABA_AR tiplerine özgü bu farklılaşmış fizyolojik görevlerin gittikçe daha belirgin hale gelmesi, ilgili reseptör tiplerine yönelik özel ilaçların geliştirilebilmesi ile ilgili ihtimalleri de akla getirmektedir. Zira, son yıllarda omurgalı, omurgasız ve prokaryot kaynaklardan elde edilen Cys-loop reseptör ailesine ait reseptörlerin X-isini kristal yapılarına dair yeni verilerin birikmesiyle çeşitli nöropatolojilere yönelik olarak sinaptik ve ekstrasinaptik GABA_AR tiplerini seçici olarak hedefleyebilen yeni ilaçların geliştirilmesi mümkün olabilecektir.

Anahtar Kelimeler: GABA (A) reseptörleri, GABA, fazik inhibasyon, tonik inhibition, gama2 (γ 2), delta (δ), Cys-loop reseptörleri, sinaptik, ekstrasinaptik

1. Introduction

Neural circuits mediate brain information processing by the integration of excitatory and inhibitory signals generated by neurotransmitters like Glutamate and GABA respectively. The actions of GABA is mediated by different receptors, including gamma-aminobutyric acid type A receptors (GABAARs) which play a significant role by means of its spatiotemporal activity during brain information processing (Koch et al., 1983). Belonging to family of "Cys-loop receptors", GABAARs are heteropentameric chloride

channels responsible for the inhibitory transmission in the vertebrate central nervous system (reviewed in Lester et al., 2004; Unwin, 2005; Sine & Engel, 2006). These GABA-gated heteropentameric channels are permeable to HCO₃⁻ and Cl⁻ ions (reviewed by Sieghart and Sperk 2002). Depending on the intracellular Cl⁻ concentration, GABAAR activation can lead to Cl⁻ influx or efflux. In adult neurons, upon activation of the receptor by GABA binding, Cl⁻ usually moves into the cell, causing a strong inhibitory hyperpolarization (Kaila et al., 1997; Rivera, et al., 2005).

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1.1. The structure of the GABAARs

Initially, the atomic structure of a GABAAR subunit complex was determined by the studies based on the snail acetylcholine receptor binding protein (AChBP) and muscle nicotinic acetylcholine receptor from the electric organ of the Torpedo ray fish (Brejc et al., 2001; Cromer et al., 2002; Ernst et al., 2003; Unwin, 2003, 2005; Sine and Engel, 2006). Recently, the crystallized structure of homeric $\beta 3$ subunit containing GABAAR (GABAAR- $\beta 3$ cryst) at 3\AA resolution has been reported (Miller and Aricescu, 2014). This study provides a direct overview for the receptor structure for the first time (Figure 1) which confirms the characteristic elements of eukaryotic Cys-loop receptors (reviewed by Lynagh and Pless, 2014). Thus GABAARs are arranged as heteropentameric structures by which five subunits co-assemble around a central pore. Each subunit has a large extra-cellular N-terminus, four transmembrane domains (TM1-TM4), and a small extracellular C-terminal. There is a large intracellular loop between the third and fourth transmembrane domains.

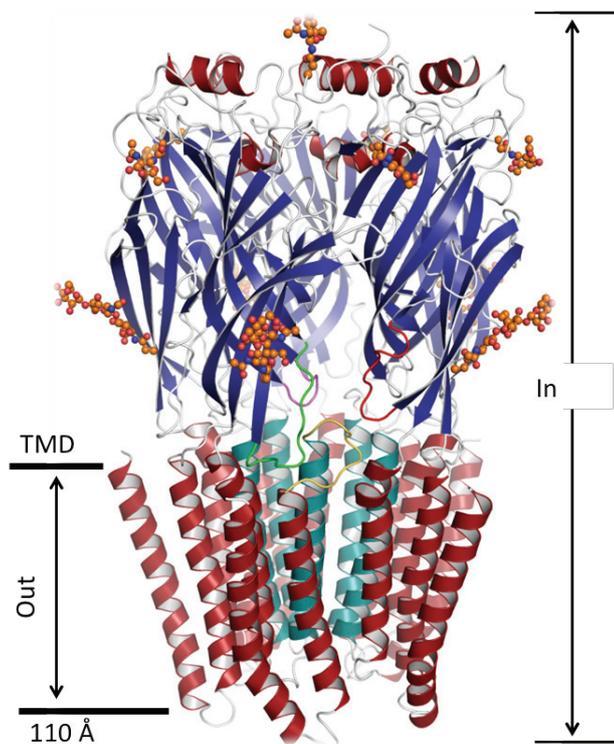


Figure 1: Crystal structure of the human GABA (A) receptor composed of $\beta 3$ homopentamer (GABA(A)R- $\beta 3$ cryst). α -helices are shown in red, excluding the pore-lining second transmembrane domain, shown in green; β -strands are shown in blue and the loops are shown as grey. Orange ball-and-stick model illustrates the N-linked glycans. (TMD: Transmembrane domain). (Reproduced from Miller & Aricescu, 2014)

1.2. Subunit composition: Tonic and Phasic Inhibition

The GABAARs are assembled from a subunit pool of 19 subunits. These subunits are named as $\alpha 1$ – $\alpha 6$, $\beta 1$ – $\beta 3$, $\gamma 1$ – $\gamma 3$, δ , ϵ , θ , η , $\rho 1$ – $\rho 3$ (Schofield et al., Korpi et al., 2002a; Rudolph and Mohler, 2006; Whiting, 2006) and expressed in age and anatomy dependent manner

(Wisden et al., 1992; Laurie et al., 1992a, b; Fritschy and Mohler, 1995; Schwarzer et al., 2001). Also cell type is a critical factor for the subunit co-assembly. It is known that the δ subunit co-assemble specifically with the $\alpha 4$ and $\alpha 6$ subunits in the forebrain (Cope et al., 2005) and cerebellum respectively (Jones et al., 1997; Peng et al., 2002). The δ represents a rare isoform of GABAAR subunits specifically located at the extrasynaptic sites (Nuser et al., 1998; Wei et al., 2003) in the dentate gyrus granule cells of hippocampus (Sun et al., 2004) and ventrobasal nucleus of the thalamus and neocortex (Chen et al., 2005; Cope et al., 2005; Glykys et al., 2007) besides to cerebellar granule cells (Jones et al., 1997; Jehlinger et al., 1998). These δ containing GABAARs receptors (δ -GABAARs) have higher affinity for GABA (Fisher and Macdonald, 1997) and mediate the tonic inhibition (Petriani et al., 2004; Farrant and Nusser, 2005), which is activated by GABA diffusing out of the synaptic cleft (Hamann, et al., 2002). Tonic inhibition is critical for the synaptic integration besides to its role in setting the threshold for the action potential generation and controlling the excitatory synaptic signalling (Hausser and Clark, 1997; Hamann, et al., 2002, Semyanov et al., 2004). It is also important to mention that tonic inhibition is not only mediated by δ -GABAARs, $\alpha 5$ containing GABAARs also mediate the tonic inhibition (Glykys, et al., 2008) which is not the focus of this study. Another GABAARs subtype is the one, that is most abundant in brain: the $\gamma 2$ containing GABAARs ($\gamma 2$ -GABAARs) are composed of $\alpha \beta \gamma 2$ subunits and mediate fast synaptic inhibition (Ernst et al., 2003). They are abundantly clustered in the postsynaptic sites besides to nonsynaptic sites (Somogyi et al., 1996; Nuser et al., 1996, 1998; Fujiyama et al., 2000, 2002, Kullman et al., 2005). Figure 2 shows the phasic and tonic inhibition mediated by synaptic and extrasynaptic GABAARs in the forebrain (Goetz et al., 2007).

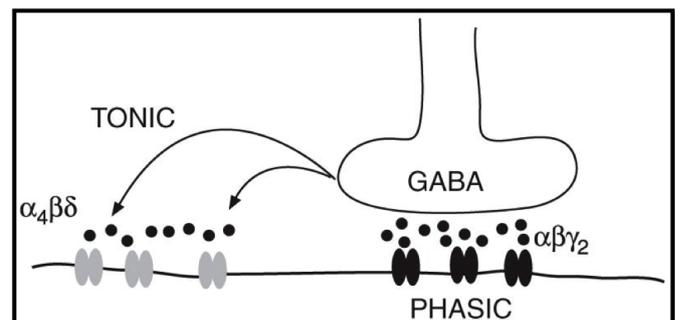


Figure 2: $\gamma 2$ -GABAARs are concentrated at the postsynaptic sites that contain α , β and $\gamma 2$ subunits (shown as black receptors at the postsynaptic site). They mediate classical fast synaptic inhibition (phasic inhibition) in response to GABA release (black dots in the synaptic cleft). δ -GABAARs contain α , β and δ subunits (shown as gray receptors at the extrasynaptic site) and mediate tonic inhibition in response to GABA spillover from the synaptic cleft (arrowed black dots). (Reproduced from Goetz et al., 2007).

1.3. Distinct physiological functions, distinct modulation

As described so far, synaptic and extrasynaptic GABAARs mediate two physiologically distinct forms of GABAergic signalling (Fisher and Macdonald, 1997; Mody and

Pearce, 2004; Farrant and Nusser, 2005). This is also well reflected in the level of their modulation. For example the $\gamma 2$ -GABAARs are sensitive to Benzodiazepines (BZ) (Prittchett et al., 1989; Wingrove et al., 1997 Sigel and Buhr, 1997). Among the specific combinations of $\gamma 2$ -GABAARs, there are also further differential modulatory effects. Sedative effects of etomidate is mediated by $\gamma 2$ -GABAARs with $\beta 2$ subunits, and anesthetic effects of etomidate is mediated by $\gamma 2$ -GABAARs with $\beta 3$ subunit (Reynolds et al., 2003). Etomidate also affects δ -GABAARs similar to allosteric modulatory effect of this anesthetic on $\gamma 2$ -GABAARs (Feng et al., 2014), but by a different kinetic mechanism than that of $\gamma 2$ -GABAARs. The etomidate potentiation of δ -GABAARs occurs by a prolonged deactivation and reduced desensitization of the receptor (Liu et al., 2015) where as the etomidate potentiation of $\gamma 2$ -GABAARs does not involve receptor desensitization (Zhong et al., 2008).

δ -GABAARs are specifically sensitive to physiologically relevant ranges of alcohol a feature that is not found in $\gamma 2$ -GABAARs (Deitrich et al., 1989; Sundstrom-Poromaa et al., 2002; Wallner et al., 2003; 2014; Liang and Olsen 2014). Another difference between the $\gamma 2$ -GABAARs and δ -GABAARs is their modulation by neurosteroids, steroids synthesized in the brain. The most widely explored neurosteroids are allopregnanolone, allotetrahydrodeoxycorticosterone, androstanediol which enhance the activity of both receptor subtypes but this enhancement is stronger for the latter as δ -GABAARs are more sensitive to neurosteroids (Wohlfarth et al., 2002; Stell et al., 2003; Carver and Reddy, 2013, Romo-Parra, et al., 2015) As a result, interaction of δ -GABAARs with neurosteroids have a critical status in terms of the potentiation of tonic inhibition and thus impacting on neuronal network excitability, seizure susceptibility, and behavior (Carver and Reddy, 2013). This has already been shown by the studies of δ subunit knock-out mice, which have a reduced inhibition and neurosteroid sensitivity (Spigelman, et al., 2003).

Recent studies in the literature continuously address the differential modulation. An interesting study not only report the differential modulation of $\gamma 2$ -GABAARs and δ -GABAARs but also further dissect δ -GABAARs in this context (Hoestgaard-Jensen, et al., 2014). The ligand 4,5,6,7-tetrahydroisothiazolo[5,4-c]pyridin-3-ol (Thio-THIP) was found to have a negligible antagonist activity at the $\alpha 4\beta 2\delta$ subtype unlike its effect on $\alpha 4\beta 1\delta$ and $\alpha 4\beta 3\delta$ among δ -GABAARs Moreover, the Thio-THIP displayed weak antagonist activity at $\alpha 1,2,5\beta 2,3\gamma 2$ containing receptors. Some other new studies report the differential modulation in terms of phasic and tonic inhibition. One study reports the potentiation of phasic and tonic currents following the inhibition of nitric oxide synthase (Gasulla and Calvo, 2015), while others suggest phosphorylation related modulation of phasic and tonic currents by CaMKII and PKC, respectively (Joo et al., 2014), or potentiation of tonic inhibition by serotonin (Jang, et al., 2015). However the subunit composition of the receptors that mediate phasic or tonic inhibition in these electrophysiological studies is not known except for the case of Gasulla and Calvo (2015), may

be. Since there are many different GABAAR subunits (see the section "Subunit composition: Tonic and Phasic Inhibition"), electrophysiological recordings of phasic and tonic inhibition, without molecular profiling, will only provide a limited understanding of receptor subtype specific modulation. The focus of the present study is only $\gamma 2$ -GABAARs and δ -GABAARs, which mediate phasic and tonic inhibition. On the other hand there is another GABAAR subtype that mediate tonic inhibition for example: the $\alpha 5$ containing extrasynaptic GABAARs (Prenosil et al., 2006; Glyks, et al., 2008). Thus, the studies of electrophysiological recording of phasic and tonic inhibition and their modulation requires further molecular investigation in this context. Nevertheless, accumulating data address the modulation of δ -GABAARs by kinase A and C (phosphorylation) and regulation of tonic inhibition by G-protein coupled receptors (see Connelly et al., 2013 for a detailed review). Modulation of $\gamma 2$ -GABAARs by phosphorylation is already a better explored subject (Kittler and Moss, 2003) and the current emerging data about the modulation of δ -GABAARs by phosphorylation as well as by other ligands described so far will add a new dimension for the distinct modulation of phasic and tonic inhibition mediated by these receptor subtypes for their significance as potential targets of subtype selective drugs.

2. Conclusion

Assembled from a large subunit pool, the rich molecular, cellular and functional diversity of GABAARs is well known (Seeburg et al., 1990; Mody and Pierce, 2004). This diversity manifests itself in the form of distinct physiological functions. Phasic and tonic inhibition mediated by synaptic and extrasynaptic GABAARs is one example of this phenomenon. What is new in the agenda is the emerging data: Studies from X-ray crystal structures of ligand-gated ion channels from prokaryotic and invertebrate organisms (Lynagh and Pless, 2014), together with the crystal structure of human GABAAR (Miller and Aricescu, 2014) and serotonin receptor (Hassaine et al., 2014) accumulate. These accumulating data will trigger the studies of molecular dynamics and homology modeling which will boost the development of new drugs for selective modulation of the GABA receptor subtypes for the treatment of various neuropathological states such as epilepsy (Rogawski et al., 2013), premenstrual dysphoric disorder (Staley and Scharmann, 2005) or alcohol use disorders (Liang and Olsen, 2014).

References

- Brejck, K., van Dijk, W.J., Klaassen, R.V., Schuurmans, M., van Der Oost, J., Smit, A.B., Sixma, T.K. (2001) Crystal structure of an ACh-binding protein reveals the ligand-binding domain of nicotinic receptors. *Nature*, 411:269-276.
- Carver C.M., Reddy D.S. (2013) Neurosteroid interactions with synaptic and extrasynaptic GABA(A) receptors: regulation of subunit plasticity, phasic and tonic inhibition, and neuronal network excitability. *Psychopharmacology (Berl)*, 230(2):151-88.
- Connelly W.M., Errington A.C., Di Giovanni G., Crunelli V. (2013) Metabotropic regulation of extrasynaptic GABAA receptors. *Front Neural Circuits*, 7:171.

- Cope, D.W., Hughes, S.W., Crunelli, V. (2005) GABAA receptor-mediated tonic inhibition in thalamic neurons. *J Neurosci.*, 25:11553-11563.
- Cromer, B.A., Morton, C.J., Parker, M.W. (2002) Anxiety over GABAA receptor structure relieved by AChBP. *Trends Biochem Sci.*, 27:280-287.
- Deitrich RA, Dunwiddie TV, Harris RA, Erwin VG. (1989) Mechanism of action of ethanol: initial central nervous system actions. *Pharmacol Rev. Dec;41(4):489-537.*
- Ernst, M., Brauchart, D., Boresch, S., Sieghart, W. (2003) Comparative modeling of GABAA receptors: limits, insights, future developments. *Neuroscience*, 119:933-943.
- Farrant, M., Nusser, Z. (2005) Variations on an inhibitory theme: phasic and tonic activation of GABAA receptors. *Nat Rev Neurosci.*, 6:215-229.
- Fisher, J.L., Macdonald, R.L. (1997) Single channel properties of recombinant GABAA receptors containing $\alpha 2$ or $\alpha 3$ subtypes expressed with $\alpha 1$ and $\alpha 3$ subtypes in mouse L929 cells. *J Physiol.*, 505:283-297.
- Fujiyama, F., Fritschy, J.M., Stephenson, F.A., Bolam, J.P. (2000) Synaptic localization of GABAA receptor subunits in the striatum of the rat. *J Comp Neurol.*, 416:158-172.
- Fujiyama, F., Stephenson, F.A., Bolam, J.P. (2002) Synaptic localization of GABAA receptor subunits in the substantia nigra of the rat: effects of quinolinic acid lesions of the striatum. *Eur J Neurosci.*, 15:1961-1975.
- Gasulla J, Calvo DJ. (2015) Enhancement of tonic and phasic GABAergic currents following nitric oxide synthase inhibition in hippocampal CA1 pyramidal neurons. *Neurosci Lett.* 90:29-34.
- Glykys J., Mann E.O., Mody I. (2008) Which GABA(A) receptor subunits are necessary for tonic inhibition in the hippocampus? *J Neurosci.* 28(6):1421-6.
- Goetz, T., Arslan, A., Wisden, W., Wulff, P., (2007) GABAA receptor structure and function in the basal ganglia. *Prog. Brain Res.* 160:21-41.
- Hamann, M., Rossi, D.J., Attwell, D., (2002) Tonic and spillover inhibition of granule cells control information flow through cerebellar cortex. *Neuron* 33(4): 625-33.
- Hassaine G, Deluz C, Grasso L, Wyss R, Tol MB, Hovius R, Graff A, Stahlberg H, Tomizaki T, Desmyter A, Moreau C, Li XD, Poitevin F, Vogel H, Nury H. (2014) X-ray structure of the mouse serotonin 5-HT3 receptor. *Nature*, 21;512(7514):276-81. doi: 10.1038/nature13552.
- Hausser, M. and Clark, B.A. (1997) Tonic synaptic inhibition modulates neuronal output pattern and spatiotemporal synaptic integration *Neuron* 19 (3) 665-78.
- Hoestgaard-Jensen K., Dalby N.O., Krall J., Hammer H., Krogsgaard-Larsen P., Frølund B., Jensen A.A. (2014) Probing $\alpha 4\beta\delta$ GABAA receptor heterogeneity: differential regional effects of a functionally selective $\alpha 4\beta 1\delta/\alpha 4\beta 3\delta$ receptor agonist on tonic and phasic inhibition in rat brain. *J Neurosci.* 34(49):16256-72.
- Jang HJ, Cho KH, Joo K, Kim MJ, Rhie DJ. (2015) Differential modulation of phasic and tonic inhibition underlies serotonergic suppression of long-term potentiation in the rat visual cortex. *Neuroscience*, 301:351-362. doi:10.1016/j.neuroscience.2015.06.018.
- Jones, A., Korpi, E.R., McKernan, R.M., Pelz, R., Nusser, Z., Makela, R., Mellor, J.R., Pollard, S., Bahn, S., Stephenson, F.A., Randall, A.D., Sieghart, W., Somogyi, P., Smith, A.J., Wisden, W. (1997) Ligand-gated ion channel subunit partnerships: GABAA receptor $\alpha 6$ subunit gene inactivation inhibits delta subunit expression. *J Neurosci.*, 17:1350-1362.
- Joo K, Yoon SH, Rhie DJ, Jang HJ. (2014) Phasic and Tonic Inhibition are Maintained Respectively by CaMKII and PKA in the Rat Visual Cortex. *Korean J Physiol Pharmacol.*, 18(6):517-24.
- Kaila, K., Lamsa, K., Smirnov, S., Taira, T., Voipio, J. (1997) Long-lasting GABA-mediated depolarization evoked by high-frequency stimulation in pyramidal neurons of rat hippocampal slice is attributable to a network-driven, bicarbonate-dependent K⁺ transient. *J Neurosci.*, 17:7662-7672.
- Kittler J.T., Moss S.J. (2003) Modulation of GABAA receptor activity by phosphorylation and receptor trafficking: implications for the efficacy of synaptic inhibition. *Curr Opin Neurobiol.* 13(3):341-7.
- Koch, C., Poggio, T., Torre, V. (1983) Nonlinear interactions in a dendritic tree: localization, timing, and role in information processing. *Proc Natl Acad Sci* 80: 2799-2802
- Kullmann DM, Ruiz A, Rusakov DM, Scott R, Semyanov A, Walker MC. (2005) Presynaptic, extrasynaptic and axonal GABAA receptors in the CNS: where and why? *Prog Biophys Mol Biol.* 87(1):33-46.
- Laurie, D.J., Seeburg, P.H., Wisden, W. (1992a) The distribution of 13 GABAA receptor subunit mRNAs in the rat brain. II. Olfactory bulb and cerebellum. *J Neurosci.*, 12:1063-1076.
- Laurie DJ, Wisden W, Seeburg PH. (1992b) The distribution of thirteen GABAA receptor subunit mRNAs in the rat brain. III. Embryonic and postnatal development. *J Neurosci.*, 12:4151-4172.
- Lester, H.A., Dibas, M.I., Dahan, D.S., Leite, J.F., Dougherty, D.A. (2004) Cys-loop receptors: new twists and turns. *Trends in Neurosciences*, 27:329-336.
- Liang J, Olsen RW. (2014) Alcohol use disorders and current pharmacological therapies: the role of GABA(A) receptors. *Acta Pharmacol Sin.* 2014 Aug;35(8):981-93. doi: 10.1038/aps.2014.50.
- Liu K, Jounaidi Y, Forman SA, Feng HJ. (2015) Etomidate uniquely modulates the desensitization of recombinant $\alpha 1\beta 3\delta$ GABAA receptors. *Neuroscience*, 300:307-13.
- Lynagh T, Pless, S.A., Principles of agonist recognition in Cys-loop receptors. *Front. Physiol.* 2014 Apr 24; 5:160. doi: 10.3389/fphys.2014.00160. eCollection 2014.
- Miller PS, Aricescu AR. (2014) Crystal structure of a human GABAA receptor. *Nature.* 2014 Aug 21;512(7514):270-5. doi: 10.1038/nature13293. Epub 2014 Jun 8.
- Mody, I., Pearce, R.A. (2004) Diversity of inhibitory neurotransmission through GABAA receptors. *Trends Neurosci.*, 27:569-575.
- Nusser, Z., Sieghart, W., Benke, D., Fritschy, J.M., Somogyi, P. (1996) Differential synaptic localization of two major γ -aminobutyric acid type A receptor subunits on hippocampal pyramidal cells. *Proc Natl Acad Sci U S A.* Oct 15;93(21):11939-44.
- Nusser, Z., Sieghart, W., Somogyi, P. (1998) Segregation of different GABAA receptors to synaptic and extrasynaptic membranes of cerebellar granule cells. *J Neurosci.*, 18:1693-1703.
- Peng, Z., Hauer, B., Mihalek, R.M., Homanics, G.E., Sieghart, W., Olsen, R.W., Houser, C.R. (2002) GABAA receptor changes in δ subunit-deficient mice: altered expression of $\alpha 4$ and $\alpha 2$ subunits in the forebrain. *J Comp Neurol.*, 446:179-197.
- Petrini EM., Marchionni, I., Zacchi P., Sieghart, W. (2004) Clustering of Extrasynaptic GABAA receptors Modulates Tonic Inhibition in Cultured Hippocampal Neurons. *J. Biol. Chem.*, 279 (44): 45833-43.
- Prenosil GA, Schneider Gasser EM, Rudolph U, Keist R, Fritschy JM, Vogt KE. (2006) Specific subtypes of GABAA receptors mediate phasic and tonic forms of inhibition in hippocampal pyramidal neurons. *J Neurophysiol.* 96(2):846-57.
- Pritchett, D.B., Sontheimer, H., Shivers, B.D., Ymer, S., Kettenmann, H., Schofield, P.R., Seeburg, P. (1989) Importance of a novel GABAA receptor subunit for benzodiazepine pharmacology, *Nature* 338: 582-585.
- Reynolds, D.S., Rosahl, T.W., Cirone, J., O'Meara, G.F., Haythornthwaite, A., Newman, R.J., Myers, J., Sur, C., Howell, O., Rutter, A.R., Atack, J., Macaulay, A.J., Hadingham, K.L., Hutson, P.H., Bellelli, D., Lambert, J.J., Dawson, G.R., McKernan, R., Whiting, P.J., Wafford, K.A. (2003) Sedation and anesthesia mediated by distinct GABAA receptor isoforms. *J Neurosci.*, 23:8608-8617.
- Rivera, C., Voipio, J., Kaila, K. (2005). Two developmental switches in GABAergic signalling: the K⁺-Cl⁻ cotransporter KCC2 and carbonic anhydrase CAVII. *J Physiol.*, 562:27-36.
- Rogawski MA, Loya CM, Reddy K, Zolkowska D, Lossin C. Neuroactive steroids for the treatment of status epilepticus. *Epilepsia.* 2013 Sep;54 Suppl 6:93-8. doi: 10.1111/epi.12289.
- Romo-Parra H, Blaesse P, Sosulina L, Pape HC. (2015) Neurosteroids increase tonic GABAergic inhibition in the lateral section of the central amygdala in mice. *J Neurophysiol.* 113(9):3421-31.
- Seeburg, P.H., Wisden, W., Verdoorn, T.A., Pritchett, D.B., Werner, P., Herb, A., Luddens, H., Sprengel, R., Sakmann, B. (1990) The GABAA receptor family: molecular and functional diversity. *Cold Spring Harb Symp Quant Biol.*, 55:29-40.
- Semyanov, A., Walker, M.C., Kullmann, D.M., Silver, R.A. (2004) Tonic active GABAA receptors: modulating gain and maintaining the tone. *Trends Neurosci.*, 27:262-269.7.
- Sieghart, W., Sperk, G. (2002) Subunit composition, distribution and function of GABAA receptor subtypes. *Curr Top Med Chem.*, 2:795-816.
- Sigel, E., Buhr, A. (1997) The benzodiazepine binding site of GABAA receptors. *Trends Pharmacol Sci.*, 18:425-429.
- Sine, S.M., Engel, A.G. (2006) Recent advances in Cys-loop receptor structure and function. *Nature* 440: 448-455
- Somogyi, P., Fritschy, J.M., Benke, D., Roberts, J.D., Sieghart, W. (1996) The $\alpha 2$ subunit of the GABAA receptor is concentrated in synaptic junctions containing the $\alpha 1$ and $\alpha 2/3$ subunits in hippocampus, cerebellum and globus pallidus. *Neuropharmacology*, 35:1425-1444.
- Sperk, G., Schwarzer, C., Tsunashima, K., Fuchs, K., Sieghart, W. (1997) GABAA receptor subunits in the rat hippocampus I: immunocytochemical distribution of 13 subunits *Neuroscience*, 80(4): 987-1000.

Spigelman, I., Li, Z., Liang, J., Cagetti, E., Samzadeh, S., Mihalek, R.M., Homanics, G.E., Olsen, R.W. (2003) Reduced inhibition and sensitivity to neurosteroids in hippocampus of mice lacking the GABAA receptor delta subunit. *J. Neurophysiol.* 90(2):903-10

Staley, K., Scharfmann, H. (2005) A woman's prerogative. *Nature Neuroscience*, 8:697-699.

Stell, B.M., Brickley, S.G., Tang, C.Y., Farrant, M., Mody, I. (2003) Neuroactive steroids reduce neuronal excitability by selectively enhancing tonic inhibition mediated by δ subunit-containing GABAA receptors. *Proc Natl Acad Sci USA*, 100:14439-14444.

Sun, C., Sieghart, W., Kapur, J. (2004), Distribution of $\alpha 1$, $\alpha 4$, $\alpha 2$, and δ subunits of GABAA receptors in hippocampal granule cells. *Brain Research* 1029: 207- 216

Sundstrom-Poromaa, I., Smith, D.H., Gong, Q.H., Sabado, T.N., Li, X., Light, A., Wiedmann, M., Williams, K., Smith, S.S. (2002) Hormonally regulated $\alpha(4)\beta(2)\delta$ GABAA receptors are a target for alcohol. *Nat Neurosci.*, 5:721-722.

Unwin, N. (2003) Structure and action of the nicotinic acetylcholine receptor explored by electron microscopy. *FEBS Lett.*, 555:91-95.

Unwin, N. (2005) Refined structure of the nicotinic acetylcholine receptor at 4 Å resolution. *J. Mol. Biol.*, 346:967-989.7.

Wallner, M., Hanchar, H.J., Olsen, R.W. (2003) Ethanol enhances $\alpha 4 \beta 3 \delta$ and $\alpha 6 \beta 3 \delta$ gammaaminobutyric acid type A (GABAA) receptors at low concentrations known to affect humans. *Proc Natl Acad Sci U S A*, 100:15218-15223.

Wallner M, Hanchar HJ, Olsen RW. (2014) Alcohol selectivity of $\beta 3$ -containing GABAA receptors: evidence for a unique extracellular alcohol/imidazobenzodiazepine Ro15-4513 binding site at the $\alpha + \beta$ - subunit interface in $\alpha \beta 3 \delta$ GABAA receptors. *Neurochem Res.* 2014 Jun;39(6):1118-26.

Wei, W., Zhang, N., Peng, Z., Houser, C.R., Mody, I. (2003) Perisynaptic localization of delta subunit-containing GABAA receptors and their activation by GABA spillover in the mouse dentate gyrus. *J Neurosci.*, 23: 10650-10661.

Wingrove, P.B., Thompson, S.A., Wafford, K.A., Whiting, P.J. (1997) Key amino acids in the γ subunit of the γ -aminobutyric acid A receptor that determine ligand binding and modulation at the benzodiazepine site. *Mol Pharmacol.*, 52: 874-881.

Wisden, W., Laurie, D.J., Monyer, H., Seeburg, P.H. (1992) The distribution of 13 GABAA receptor subunit mRNAs in the rat brain I: Telencephalon, diencephalon, mesencephalon. *J Neurosci.*, 12:1040-1062.

Wisden, W., Cope, D., Klausberger, T., Hauer, B., Sinkkonen, S.T., Tretter, V., Lujan, R., Jones, A., Korpi, E.R., Mody, I., Sieghart, W., Somogyi, P. (2002) Ectopic expression of the GABAA receptor $\alpha 6$ subunit in hippocampal pyramidal neurons produces extrasynaptic receptors and an increased tonic inhibition. *Neuropharmacology.*, 43:530-549.

Wohlfarth, K.M., Bianchi, M.T., Macdonald, R.L. (2002) Enhanced neurosteroid potentiation of ternary GABAA receptors containing the δ subunit. *J Neurosci.*, 22:1541-1549.

Zhong H, Rüsç D, Forman SA. Photo-activated azi-etomidate, a general anesthetic photolabel, irreversibly enhances gating and desensitization of gamma-aminobutyric acid type A receptors. *Anesthesiology.* 2008 Jan;108(1):103-12.